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Article

Super-Earths, M Dwarfs, and Photosynthetic Organisms: Habitability in the Lab

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Abstract: In a few years, space telescopes will investigate our Galaxy to detect evidence of life, mainly by observing rocky planets. In the last decade, the observation of exoplanet atmospheres and the theoretical works on biosignature gasses have experienced a considerable acceleration. The most attractive feature of the realm of exoplanets is that 40% of M dwarfs host super-Earths with a minimum mass between 1 and 30 Earth masses, orbital periods shorter than 50 days, and radii between those of the Earth and Neptune ($1\text{--}3.8 R_{\oplus}$). Moreover, the recent finding of cyanobacteria able to use far-red (FR) light for oxygenic photosynthesis due to the synthesis of chlorophylls *d* and *f*, extending in vivo light absorption up to 750 nm, suggests the possibility of exotic photosynthesis in planets around M dwarfs. Using innovative laboratory instrumentation, we exposed different cyanobacteria to an M dwarf star simulated irradiation, comparing their responses to those under solar and FR simulated lights. As expected, in FR light, only the cyanobacteria able to synthesize chlorophyll *d* and *f* could grow. Surprisingly, all strains, both able or unable to use FR light, grew and photosynthesized under the M dwarf generated spectrum in a similar way to the solar light and much more efficiently than under the FR one. Our findings highlight the importance of simulating both the visible and FR light components of an M dwarf spectrum to correctly evaluate the photosynthetic performances of oxygenic organisms exposed under such an exotic light condition.

Keywords: astrobiology; M stars; super-Earths; photosynthesis



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1. Introduction

The considerable number of new worlds discovered so far is pushing scientists to provide evidence of life on other planets. The diversity in kind, composition, masses, and radii of these new worlds is so vast that almost all possible mass values are covered in continuity from Mars ($0.11 M_{\oplus}$) up to super-Jupiters ($>10 M_J$). Among all the planetary hosts, low mass stars, mainly M spectral type stars, are the main targets of the extrasolar planet surveys due to both their high density in the Galaxy and their small radii that provide higher amplitude transit signals than solar-like stars [1,2]. Indeed, the most attractive characteristic of these systems is that 40% of M stars host super-Earths with a minimum mass between about 1 and 30 Earth masses, orbital periods shorter than 50 days, and radii between those of the Earth and Neptune ($1\text{--}3.8 R_{\oplus}$). Due to these high occurrence rates, super-Earths ($1\text{--}10 M_{\oplus}$) represent the most common type of components of planetary

systems in the Galaxy [3]. Even more striking, the frequency of super-Earths found in the habitable zone (HZ) of M dwarfs (with a period between 10 and 100 days) is about 50% [4,5]. These results renew, with higher and more interdisciplinary efforts, the search for life as an astrophysical problem. In this framework, it is critical to determine the types of biosignatures (based on the so-far-recognized life signatures) for when designing the next generation of ground- and space-based instruments that will observe these planets at both high spectral and spatial resolutions (e.g., Reference [6–13]). Among all the biosignatures, oxygen seems to be the most prominent signature that can reveal the existence of life on other planets [14]. In this sense, it is our most robust and the most studied biosignature [15]. Its presence, together with other gases, like CH₄ or N₂O, is the signal of a thermodynamic disequilibrium that, for a long time, has been considered as compelling evidence for life (e.g., Reference [16]). Since the closure of Seager & Bains (2015) [17], many arguments countered this concept because using the thermodynamic disequilibrium as a biosignature cannot be easily generalized. Other authors have also argued that oxygen is not a suitable bioindicator due to photochemical reactions that have abiotic O₂ as a byproduct (e.g., H₂O and CO₂ photoionization). A detailed discussion of several possible false positives is presented in Harman et al. (2015) [18].

Today, in the atmosphere of Earth, oxygen is highly abundant (21% by volume) due to the oxygenic photosynthesis of plants, algae, and cyanobacteria, the presence of which can be detected by remote observations, not only for the presence of O₂ in the atmosphere of the planet but also by discerning the red edge. The red edge is a feature associated with the high reflectance of photosynthetic organisms at near infra-red (NIR) in contrast with the absorption by chlorophyll in wavelengths shorter than about 700 nm. This phenomenon emerges due to the scattering of light at the interfaces between the cell walls and the air space inside the organism [19].

In recent years, biologists have found species of cyanobacteria able to use far-red (FR) light for oxygenic photosynthesis due to the synthesis of chlorophylls *d* and *f* [20], extending in vivo light absorption up to 750 nm, suggesting the possibility of exotic photosynthesis in planets around M stars. So far, a number of works have discussed the possibility of emerging oxygenic photosynthesis on a planet in the HZ of an M star (e.g., Reference [21–23]) under favorable conditions.

Considering all these favorable observational and theoretical circumstances, it is important to assess, in an experimental way, the consequences of oxygenic photosynthesis on planets orbiting in the HZ of M stars. In this paper, we present the laboratory set up and experiments conducted with the aim of understanding the performance of photosynthetic organisms exposed to conditions similar to that of an Earth-like planet in the HZ of an M star. In particular, the laboratory set up simulates the exoplanetary surface temperature and radiations. In these experiments, we analyzed the growth and the photosynthetic efficiency of several cyanobacteria with different photosynthetic behaviors, based on chlorophyll fluorescence measurements. In particular, we selected a cyanobacterium unable to utilize FR light for carrying out oxygenic photosynthesis, two species able to exploit it and a species both able to use FR light and perform the so-called Chromatic Acclimation (CA), changing its color depending on the incident light spectrum.

This paper is organized as follows: In Section 2, we describe the approach to the problem and the experimental plan. In Section 3, the experimental set up and its validation is described; in Section 4, we discuss the microorganisms we used and report the results of the experiment. Section 5 is allocated to the discussion and the conclusion.

2. Experimental Aims

Our research plan aim was analyzing the changes caused by the presence of photosynthetic bacteria to the chemical composition of a simulated, secondary atmosphere of a super-Earth inside the HZ of an M star. This expression of intents delimits the environmental parameters (pressure, temperature, irradiation pattern, initial atmospheric chemical composition) we want to simulate in the laboratory. The description of another version of

this experiment is detailed in Reference [24]. Here, we describe an entirely new evolution of the experiment, putting it into context.

2.1. The Astrophysical Context

M dwarfs are faint hydrogen-burning stars with masses ranging between $0.6\text{--}0.075 M_{\odot}$ and very low photospheric temperatures (3870 K for an M0 V and 2320 K for an M9 V star [25]). Their spectra are characterized by maximum emission at longer wavelengths than those of the Sun and molecular band absorptions that deplete the emitted visible flux (see Figure 1). In the visible, the Sun (G2 V) has an absolute magnitude of about 4.8, while the M stars are about 4 orders of magnitude fainter than the sun (the absolute visual magnitude of an M star ranges between 9 for M0 V to 20 mag for M9 V [25]).

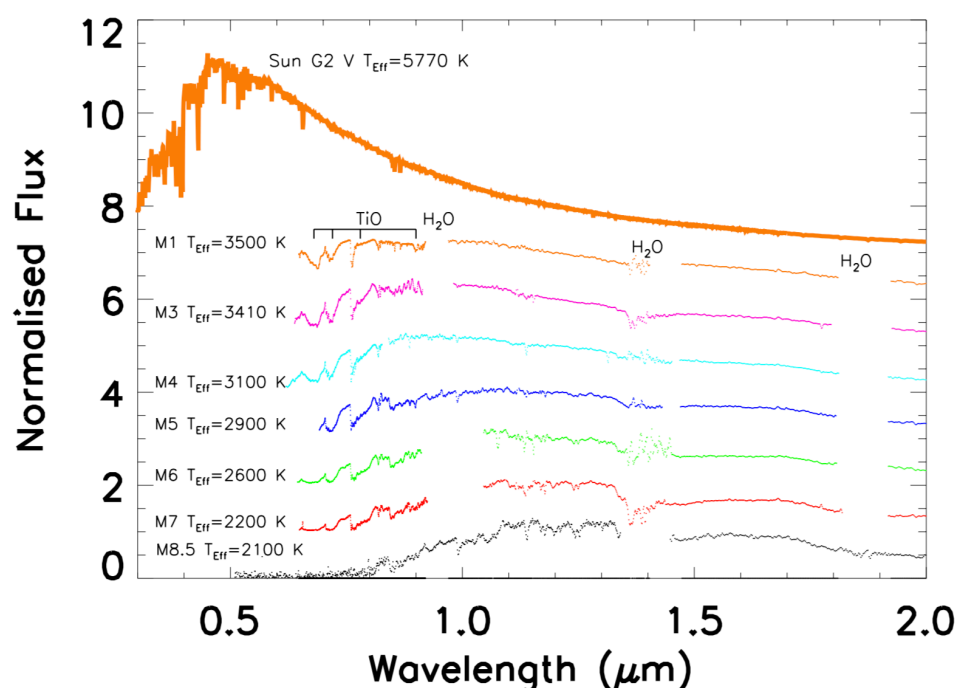


Figure 1. Examples of various M-stars spectra [26], from higher to lower photospheric temperatures. The main molecular bands depleting the visible and near infra-red (NIR) flux are also indicated. The spectral features of M-stars are indicated in Reference [27]. The upper part of the figure shows the spectrum of the Sun for comparison. The Sun and M star spectra have been normalized to the flux they emit at $1.2 \mu\text{m}$ and offset.

In other words, M dwarfs are very different from the Sun in both luminosity and spectral distribution. Luminosity affects the position of the HZ around the star, whereas spectral distribution influences the number of photons available in the PAR (Photochemically Active Radiation) wavelength range ($400 \leq \lambda \leq 700 \text{ nm}$), challenging the possibility of photosynthesis on the surface of a rocky planet orbiting in the HZ of this kind of star.

Not only the wavelength range of the spectral peak, but also the qualitative shape of the spectrum and how it interacts with an HZ planetary atmosphere, vary significantly across the M star mass range. In fact, due to the faintness of M stars, the HZ is as close as $\sim 0.30 \text{ au}$ or even closer to the star [5,28,29]. Hence, the planet orbiting inside the HZ results as tidally locked and becomes a synchronous rotator with 1:1 spin-orbit resonance (e.g., Reference [28]) or higher (3:2) (e.g., Reference [30]). Another possible consequence to consider is whether there is sufficient flux in the wavelength range used for photosynthesis [31]. Heath et al. (1999) [32] determined that the PAR from M stars would be lower than the average terrestrial value by about one order of magnitude.

In past years, several authors have also argued that terrestrial planets within the HZs of M dwarfs may not be habitable. The main reasons range from the possible deficiency of volatiles in the planetary atmosphere [33], to the scarce water delivery during the planet evolution [34], or the loss of planetary water during the pre-main sequence due to the higher luminosity of the protostar in that evolutionary phase [35]. The latter seems a showstopper indeed. In any case, Tian and Ida (2015) [36] showed that the content of water in Earth-like planets orbiting low mass stars could be rare, but dune and deep ocean worlds may be common.

Nevertheless, many authors are optimistic asserters that the oxygenic photosynthesis can take place on a super-Earth surface also under these conditions. Recent studies on possible water loss in the atmosphere of planets orbiting very cool stars, like Trappist-1 d, show that these planets may still have retained enough water to support surface habitability [37,38]. Furthermore, previous work on the photosynthetic mechanisms and spectral energy requirements elucidated that photosynthesis can still occur in harsh and photon-limited environments (e.g., Reference [21–23,32,39]).

Another possible showstopper discussed in several works is the obstacle caused by the stellar activity. M dwarfs, by nature, are characterized by their high stellar activity. These stars can significantly change their activity depending on their evolutionary stage. During a quarter of their early life, M dwarfs release high amounts of XUV through flares and chromospheric activity [23], while quiescent stars emit little UV radiation and have no flares [40]. Planets orbiting around M dwarfs often receive high doses of XUV radiation during stellar flares, with fluxes that can increase ten to a hundred times and occur 10–15 times per day. These events rapidly change the radiation environment on the surface [41–43] and possibly erode the ozone shield, as well as parts of the atmosphere. However, some researchers point out that these planets could remain habitable despite these issues [41,44,45]. The presence of a strong magnetic field or a thick atmosphere [46,47] could avoid planetary atmosphere erosion. On these planets, possible organisms could develop UV protecting pigments and DNA repair mechanisms, similar to Earth, or thrive in subsurface niches [48] and underwater, where radiation is less intense. This kind of life, of course, would not be detectable remotely. A UV-protective mechanism that could be detected remotely is the photo-protective bioluminescence, where proteins shield the organisms from dangerous UV radiations, emitting the energy at longer, detectable wavelengths. Moreover, it would allow organisms to live on the surface of planets orbiting active stars and is a mechanism already at play on Earth for some species of corals [45]. The flaring activity of M dwarfs, in particular, could even be beneficial for oxygenic photosynthesis, if the XUV portion is adequately shielded by the atmosphere. They could indeed enhance the effectiveness of oxygenic photosynthesis due to the additional flux in the PAR that they can generate [49], even if they are not thought to allow an Earth-like biosphere in planets orbiting the HZ of the majority of the M dwarfs known [13].

From an observational point of view, *Kepler* found that Earth-sized planets ($1.0\text{--}1.5 R_{\oplus}$) are common around M stars with an occurrence rate of 56% with periods shorter than 50 days. Super-Earths with radii between $1.5\text{--}2.0 R_{\oplus}$ and periods shorter than 50 days orbit M dwarfs with an occurrence rate of 46% [50]. Similar high occurrence rates are reported by the radial velocity surveys [3]. Notable examples are the Trappist-1 system with seven super-Earths [51] and Proxima Cen b [52] orbiting two M stars.

These planets are predicted to have large surface gravities (25 ms^{-2} for $5 M_{\oplus}$ (Reference [53] and References therein)) and are likely to exist within a wide range of atmospheres: some of them could be able to retain a thick H-rich atmosphere, whereas others could have a stronger resemblance to Earth with heavier molecules in their atmospheres. A third possibility could be an atmosphere with a moderate abundance of hydrogen due to its escape and/or molecular hydrogen outgassing [53].

2.2. The Experiment Plan

The background scenario is crowded with theoretical hypotheses on the photosynthesis at work on planets orbiting M stars; however, to the knowledge of authors, no experimental work has been performed directly exposing photosynthetic organisms to a simulated M-dwarf spectrum. Here, the (unavoidable) working hypothesis is that the evolution of extra-terrestrial life converged to pigment production and photosynthetic mechanisms similar to that of terrestrial extremophiles under non-Earth conditions.

Our simple approach consists of the following steps:

- i We set up the laboratory instrumentation and selected the organisms for the tests (see Section 4.1). We built some of the laboratory tools ex-novo. We first built a light source suitable for the purpose of the experiment, simulating the star irradiation (see Section 3.1). Secondly, we built the reaction cell (see Section 3.2).
- ii Before conducting the main experiment, we performed a fiducial one considering the terrestrial environment. We irradiated the selected organisms with solar light and within a terrestrial atmosphere environment.
- iii Once we checked that the experimental set up functions well in terrestrial condition, we switched to the M star irradiation of organisms, considering a terrestrial atmosphere.
- iv Lastly, we are planning to conduct experiments using the M star light to irradiate the cyanobacteria that will be put in a modified atmosphere. The composition is defined using the 1-D model of the atmosphere of super-Earths described by Petralia et al. 2020 [54] and Alei et al. 2020 (submitted).

In this paper, we address the first three points. The fourth one will be part of future work.

3. Laboratory Set Up

The experimental set up is sketched in Figure 2. It can be conceptually split into two parts: the stellar simulator on top, composed of the illuminator and a spectrometer, with the reaction cell on the bottom, where the microorganisms used for the experiments are hosted. The entire system is isolated in a dark container and is cooled by two fans, it is equipped with an anti-condensation system, and it is monitored through a webcam. A control PC allows to operate remotely the system without interfering with the experiment.

3.1. Star Irradiation Simulator

To achieve the described experimental aim, it was necessary to have an unconventional light source. In particular, the light sources used in photosynthetic study facilities are mainly metal halide, high-pressure sodium, fluorescent, and incandescent lamps. These lamps are commercially available and mostly emit solar or close to solar radiation, with a limited capability in adjusting the color temperature and the intensity of the output radiation. In our experiment, we need a light source able to reproduce the irradiation of stars other than the Sun in a quite simple and direct way. Furthermore, it should be operable without interfering with the experiment. To achieve our goal, we designed a completely different light source using light-emitting diodes (LEDs) controlled remotely by a computer.

LEDs have also been used in the laboratory as light sources for their efficiency in plants growth [55]. For that purpose, the LED-based devices are built to illuminate the material in the red part (600–700 nm) of the PAR, while, typically, the blue part of PAR is covered by blue fluorescent lamps.

For our purposes, we need a completely different device. In fact, for Wien's law ($\lambda_{\text{Max}}T = 290 \times 10^4 \text{ nm K}$), different stars of different spectral types have the maximum of their emission at different wavelengths. In particular, while the Sun has a peak of emission at about 550 nm, an M star, which is about a factor of 2 cooler than the Sun, has its emission peak in the NIR range (at about 1000 nm). In order to appreciate the differences in the slope of the spectra of the stars of different spectral types, we need a collection of LEDs able to cover a slightly longer wavelength range than the PAR, between about 350–1000 nm.

Moreover, this device shall be able to modulate the LEDs intensity in order to mimic, as close as possible, the flux variation of stars of different spectral types.

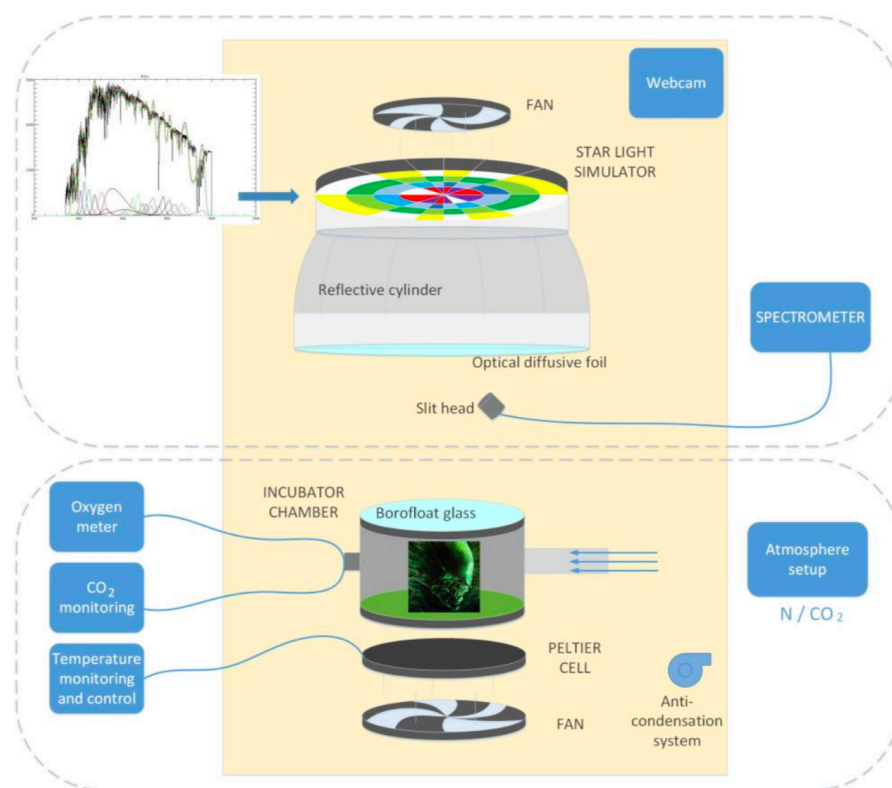


Figure 2. The instrumental set up. The upper part shows the starlight simulator (composed of the illuminator and a spectrometer); the bottom panel shows the incubator chamber, where the photosynthetic microorganisms used for the experiments are hosted.

The available LEDs allow us to consider the wavelength range between 365 nm and 940 nm covered by 25 dimmable channels (see Table 1). Because LEDs covering such wavelength range are manufactured with different technologies (from AlGaIn/InGaP to GaAs/InGaP), their emitted luminosities are also different from each other, and each channel has a different number of LEDs to achieve the required optical power at a specific wavelength. Furthermore, we added a white LED with a correlated color temperature (CCT) of 2200–2780 K to fill the spectrum in the 630 nm region. We used 312 LEDs in total, arranged in five concentric rings on which the mosaic of circuit boards is arranged in a pie-chart shape, on the surface of which the diodes have been welded [56]. Each channel is tunable enough to allow us to reproduce the radiation of stars of F, G, K, and M spectral types.

The modularity design of the board permits easy maintenance in case of damage, allowing us to remove only the problematic piece. The disposition of diodes on the board was designed to reduce the non-uniformity of the flux, due to the intrinsic light exit angle of each led. Moreover, a reflective cylinder and an optical diffusive foil were mounted to increase the uniformity. Since the thermal power of the system dominates its radiation power, the diodes are cooled by a fan set on the back of the board. A spectrometer collects the light through a slit head placed at a manually adjustable distance from the diffusive foil. The adopted spectrometer is a Component Off The Shelf (COTS) component. We selected the FLAME VIS-NIR by Ocean Optics, in which its 2040×2040 pixel detector covers the wavelength range 190–1100 nm.

Table 1. The twenty-five channels of the star irradiation simulator, together with the measured λ_{peak} and the light-emitting diode (LED) codes.

Nominal λ_{peak} (nm)	Measured $\lambda_{peak} \pm 4\%$ (nm)	LEDs Number	Tot. Luminosity @0.7A&25° (W)	LED Type
365.0	368.0	5	2.735	Engin LZI-00U600
385.0	390.0	15	2.140	Lumileds LHUV-0380-0200
405.0	404.0	10	6.030	Lumileds LHUV-0400-0500
425.0	424.0	10	8.770	Lumileds LHUV-0420-0650
447.5	450.0	10	6.980	Lumileds LXZ1-PR01
470.0	476.0	10	4.670	Lumileds LXZ1-PB01
485.0	469.0	7	5.470	Osram CRBP-HXIX-47-1
505.0	499.0	25	7.400	Lumileds LXZ1-PE01
530.0	520.0	10	2.260	Lumileds LXZ1-PM01
567.5	548.0	45	22.500	Lumileds LXZ1-PX01
590.0	604.0	20	1.530	Lumileds LXZ1-PL01
627.0	634.0	10	3.780	Lumileds LXZ1-PD01
655.0	665.0	10	4.150	Lumileds LXZ1-PA01
680.0	689.0	14	2.814	Roithner SMB1N-680
700.0	708.0	10	2.070	Roithner SMB1N-700
720.0	727.0	11	2.220	Roithner SMBIN-720D
740.0	738.0	8	4.100	Engin LZI-00R300
760.0	763.0	6	2.390	Roithner SMB1N-760D
780.0	777.0	8	3.390	Roithner SMB1N-780D
810.0	807.0	8	4.060	Roithner SMB1N-810D
830.0	834.0	15	3.860	Roithner SMB1N-830N
870.0	871.0	6	3.920	Osram SFH 47155
880.0	889.0	16	4.700	Roithner SMB1N-880
940.0	972.0	9	5.820	Osram SFH 4725S
white (2200 K)	605.0	14	5.820	Lumileds 997-LXZ1-2280-5-2200

The illuminator is controlled by a custom control software [57] that, by means of a graphical user interface (GUI), allows the user to select an appropriate spectrum chosen from a spectral library. For the input spectrum, the control software calculates the intensities of the 25 channels to best fit the spectrum. In any case, through the GUI, the user has access to each channel of the illuminator setting the output flux of the channel. The set spectrum is shown in a window of the GUI. The emitted spectrum is registered by the spectrograph and is superimposed on the input spectrum. Slight differences between the two can be fixed by adjusting the luminosity of each channel. The left panel of Figure 3 shows the simulated spectrum of a solar star (light SOL; see Section 4.2), while the right panel of the same figure shows the simulated M star spectrum (light M7; see Section 4.2). In both panels the input spectrum is represented in red color, and the emitted spectrum in blue. The input spectra are smoothed (e.g., see Figure S1 in the Supplementary Materials for an M7 V star) due to the difficulties in reproducing the high resolution stellar spectra by the spectrum simulator.

3.2. The Reaction Cell

The incubator cell (see Figure 4) is a steel cylinder of 0.5 l of volume in which the light enters through a thermally resistant Borofloat glass, with over 90% transmission in 365–940 nm wavelength range. The atmosphere in the cell can be flushed to change the initial O₂, CO₂, and N₂ levels. The cell is provided with pipe fittings and connected to an array of flow meters and needle valves (each for a different input gas: N₂, O₂, CO₂) to inject atmospheres of controlled and arbitrary compositions. Once the desired mixture is flushed through the cell, the input and output valves are closed to seal the inside environment and leave it to its evolution. Water vapor will quickly reach saturation value, due to the water-based medium in the sample Petri dish. When using high carbon dioxide levels, caution should be exercised to ensure a long enough flushing time to achieve equilibrium between the gas phase and dissolved CO₂ due to its high solubility. The base of the cell and the sample are kept at a constant temperature by controlling a Peltier cell on which the cylinder is leaned. The Peltier temperature set point is always kept 2° C lower than the surrounding

environment (which its temperature is also controlled) to avoid any condensation on the upper glass window.

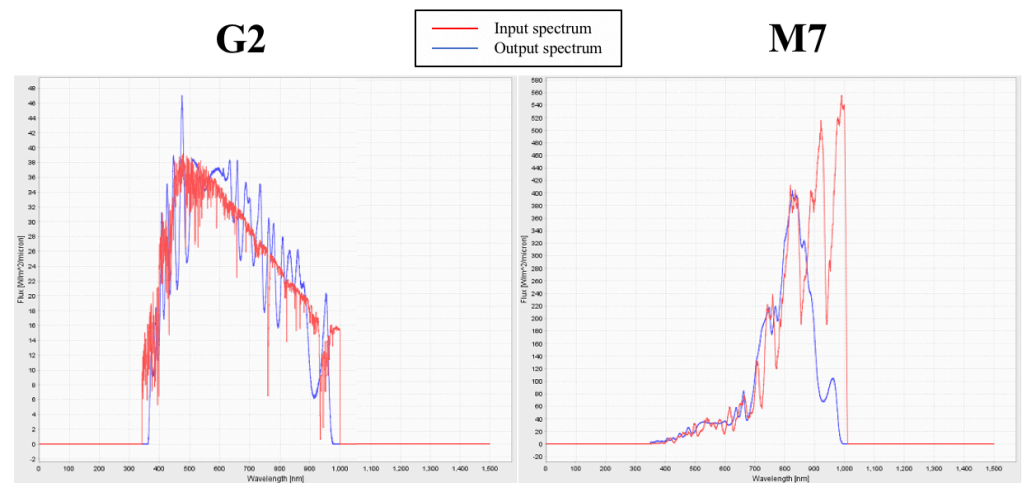


Figure 3. Left panel: G2 V solar spectrum (input) in red and the emitted light spectrum (SOL) in blue. Right panel: M7 star spectrum (input) in red and the emitted light spectrum (M7) in blue. In both panels, the spectra are plotted in $\text{W m}^{-2} \mu\text{m}^{-1}$ versus wavelength (nm).

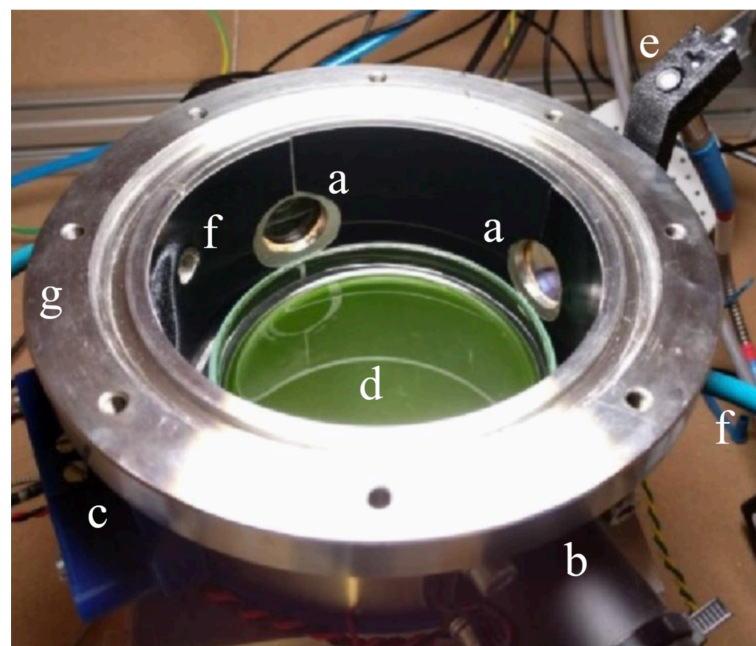


Figure 4. Reaction cell inside the Starlight simulator box. Side windows (a) provide access to the CO₂ Tunable Diode Laser Absorption Spectroscopy (TDLAS) sensing channel (b) and to the fluorescence quenching O₂ sensor (c). The reaction cell contains a Petri dish with target organisms inside (d). An optical fiber (e) provides feedback to the illuminator control spectrometer. The atmosphere inside the cell is flushed through input and output pipes (f). The flange (g) seals the cell with the top window (removed in this picture).

In the context of this work, the cell was always operating at ambient pressure and 30 °C temperature with an initial composition of 75% vol. N₂, 20% vol. O₂, and 5% vol. CO₂; this provides a high enough amount of carbon dioxide to be fixed into biomass throughout the experiment without excessively stressing the sample. Vital photosynthetic microorganisms in a liquid medium inside the cell are expected to produce oxygen. Hence, the cell is provided with a commercial fluorescence quenching oxygen sensor (Nomasense O₂

P300), while the CO₂ concentration is monitored via a custom Wavelength Modulation Spectroscopy (WMS) Tunable Diode Laser Absorption Spectroscopy (TDLAS, [58]) set up. To monitor the gas, four wedged windows of 2.5 cm are pierced and paired two by two in opposite positions on the wall of the cell. Two of the windows are used by the CO₂ sensor for TDLAS, whereas one is used for the fluorescence quenching tablet, which is remotely sensed through an optical fiber. The reaction cell underwent several modifications for reducing the systematic errors and the human interferences during the experiments. In Battistuzzi et al. (2020) [59], a complete description of the very last version of the reaction cell and the results obtained from a biological point of view is presented.

3.3. The Control Software

The starlight simulator (illuminator and spectrometer) and the incubator cell environment (gas sensors and Peltier cell) are controlled by two separate processes, running on the same computer. We wanted a stand-alone software to control the simulator because it could also be used also for other laboratory applications (e.g., photo-bioreactors, microscopy, yeast growth [60]).

3.4. Validation of the Experimental Set up

To validate the experimental set up, we positioned the cyanobacterium *Synechocystis* sp. Pasteur Culture Collection (PCC) 6803 liquid cultures into the simulator chamber with an atmosphere consisting of a mixture of gasses in the following composition: 75% of N₂, 20% of O₂, and 5% of CO₂. Eventually, we irradiated it by means of the star simulator with a solar (G2 V) spectrum with three different intensities: 30, 45, and 95 $\mu\text{mole m}^{-2} \text{s}^{-1}$ (from here, μmole is used for μmole of photons). The organisms exposed to different light regimes grew with good photosynthetic efficiency. Description of the test and of the developed method to evaluate the growth of bacteria without any interferences by the operators are fully detailed in Battistuzzi et al. (2020) [59].

4. Biological Experiment as Proof of Concept

4.1. Selected Organisms

On Earth, the rise of photosynthetic organisms transformed in billions of years the primordial planet into our beloved and green planet. A review of the fundamental processes at play in photosynthetic organisms with a set of algae and plants on Earth is presented in Ref. [23]. Among microbes, the most relevant in changing the characteristics of our planet were cyanobacteria that evolved oxygenic photosynthesis deeply influencing the Earth's atmosphere and leading to the "oxygen-breathing life" development. The recent finding of cyanobacteria able to use FR light for oxygenic photosynthesis due to the synthesis of chlorophylls *d* and *f*, extending in vivo light absorption up to 750 nm [20,61], suggests the possibility of exotic photosynthesis in planets around M stars.

We took advantage of the availability of a large selection of extremophiles, including cyanobacteria from soils, thermal spring muds and cave rocks, and cyanobacteria with UV-absorbing pigments. Extremophile cyanobacteria from environments characterized by low irradiances, rich in FR wavelengths [20], are selected for M dwarf star simulations. Some of these cyanobacteria are already known for coping with conditions not occurring in their natural environments, such as space and martian simulated conditions in low Earth orbit [62–64]. These astrobiological experiments pointed out that the limits of life have not been established well yet, and that extremophiles may have the potential to cope with the simulated environments planned in our experiment and, hence, can be a good source to identify potential astronomical biosignatures.

The selected extremophiles are expected to perform photosynthesis using the simulated M star light radiations, considering that the minimum light level for photosynthesis is about 0.01 $\mu\text{mole m}^{-2} \text{s}^{-1}$, i.e., less than 10^{-5} of the direct solar flux at Earth in the PAR wavelength range (2000 $\mu\text{mole m}^{-2} \text{s}^{-1}$) (Reference [65,66] and References therein). This implies that, even at the orbit of Pluto, light levels exceed this value by a factor of 100.

In particular, for our experiment, we chose two cyanobacteria species, *Chlorogloeopsis fritschii* PCC 6912 and *Chroococcidiopsis thermalis* PCC 7203, which are able to perform FR light photoacclimation ((FaRLiP) [20,67]), and *Synechococcus* PCC 7335, a peculiar cyanobacterium strain capable of both FaRLiP and Chromation Acclimation (CA). We compared the responses of these species with those of *Synechocystis* sp. PCC 6803, a cyanobacterium unable to activate FaRLiP or CA, hence used as control organism.

Chlorogloeopsis fritschii PCC 6912 is an organism that can thrive under various environmental conditions in terms of intensity and temperature. On Earth, its favorable habitats are thermal springs and hyper-salty lakes. The peculiarity of this strain is its ability to synthesize chlorophylls *a*, *d*, and *f*. Chlorophylls *d* and *f* are produced in a larger quantity when *Chlorogloeopsis fritschii* grows in the FR light [68].

On the other hand, *Chroococcidiopsis thermalis* PCC 7203 is a cyanobacterium isolated from a soil sample in Germany and ascribed to the species *Chroococcidiopsis thermalis* Geitler, whose type locality is Sumatra hot springs (according to Algaebase website: <https://www.algaebase.org/>). Moreover, members of the *Chroococcidiopsis* genus are widespread and can be found in freshwaters, salt waters, and hot and cold deserts [69]. Due to their capability of withstanding different and extreme conditions, *Chroococcidiopsis* strains are utilized in astrobiology studies [63]. As for *Chlorogloeopsis fritschii*, *Chroococcidiopsis thermalis* performs FaRLiP, and both grow continuously and photoautotrophically in FR light and are utilized as model organisms to study cyanobacteria acclimation mechanisms to this light source [70].

Synechococcus sp. PCC 7335 was originally isolated from a snail shell in an intertidal zone and thus adapted to changes in light regimes and hydration/dehydration. This organism is unique due to its capability to activate FaRLiP response when grown under FR light and to clearly show changing its pigmentations complementarily to the incident light spectrum, due to the so-called CA acclimation response [71].

Regarding *Synechocystis* sp. PCC 6803, it is a well-known cyanobacterium used as a model strain due to the complete sequencing of its genome. *Synechocystis* sp. PCC 6803 has been selected as a control organism. In fact, it does not possess the gene cluster that is responsible for the FaRLiP, and it does not acclimate when exposed to FR light.

4.2. Growth and Photosynthetic Efficiency

To perform the first part of the experiment (see Section 2.2), the selected cyanobacteria were grown in BG-11 medium [72] or in ASN-III medium [71], depending on the species, in both liquid and solid (with the addition of 10 g l⁻¹ of Agar) cultures. The liquid cultures were exposed to terrestrial atmospheric air in a climatic chamber maintained at a temperature between 28 and 30 °C under a continuous cool white fluorescent light of 30 µmole m⁻² s⁻¹ (L36W-840, OSRAM). Once the organisms were in the exponential growth phase, we subdivided them into spots over agarized solid medium in petri plates with BG-11 or ASN-III, to be exposed to the different light sources: solar (SOL; see Figure 3, left panel) with 20.3 µmole m⁻² s⁻¹ in the PAR and 27.4 µmole m⁻² s⁻¹ in the whole working range (380–780 nm), M7 V star (M7 see Figure 3, right panel) with 20.3 µmole m⁻² s⁻¹ in the PAR and 57.6 µmole m⁻² s⁻¹ in the whole working range, and a monochromatic flux centered at 720 nm (far-red (FR); see Figure S2 in the Supplementary Materials) with 2.3 µmole m⁻² s⁻¹ in the PAR and 20 µmole m⁻² s⁻¹ in its working range (660–780 nm) (This source of light is a small lamp built with five red LEDs SMD (Surface Mount Device) OSOLON 720). The PAR luminosity for the simulated M star is close to the values evaluated by Reference [23], incorporated in their Table 2.

For the experiment, we arranged plates with several spots of 20 µL at different cell concentrations with 0.3, 0.5, 0.7, and 1 of optical density (OD) measured at 750 nm for each organism (see Figure S3 in the Supplementary Materials). The plates were illuminated by the three different sources for 240 h and analyzed after 72 h and at the end of the experiment (see Figure S4 in the Supplementary Materials). All the experiments were performed in three biological replicates.

The top panel of Figure 5 shows the situation of the different phenotypes before and after 72 h and 240 h of irradiation with the three different light sources. An enhancement of the optical density of the spots of all the species that are greener than those at the beginning of the experiment can be observed in Figure 5. This trend is visible for both the sample irradiated by the solar light and the sample irradiated by the light with an M star simulated spectrum. The bacteria behaves differently from when they are exposed to the irradiation of the FR light as expected. In fact, it is possible to see that, unlike the other strains, the control organism (PCC 6803) does not grow under the monochromatic FR light.

These results are confirmed and quantified by the values of the F_0 incremental ratios (Figure 5, bottom panel). F_0 is the ground chlorophyll fluorescence measured with Pulse-Amplitude Modulation (PAM) on cells adapted to the dark for 20–30 min. Eventually, they are exposed to a pulse modulated light that does not trigger the photosynthetic process [73]. The chlorophyll ground fluorescence (F_0) is proportional to the increment of chlorophyll, and thus, to the number of cells in the considered spot. So, an increase of the F_0 parameter is a measure of the growth of the culture [74]. The measurements are considered reliable when the F_0 value increases linearly with the increasing OD. For this reason, we performed experiments with 4 different initial culture concentrations (OD) for each organism, and we repeated it with 3 independent biological replicates. The best initial OD meeting these conditions proved to be 0.5, 0.7, and 1 for each organism. The measurements have been performed at the beginning of the experiment ($F_{0(0H)}$) and after 72 h ($F_{0(72H)}$). We did not consider the measurement at 240 h as the signals of fluorescence with the initial PAM settings were saturated for most of the spots due to the very high cell concentrations. F_0 chlorophyll fluorescence was measured on the entire spot at 0 and 72 h, by analyzing the same area and maintaining the same acquisition setting each time [74]. The F_0 detected at the beginning ($F_{0(0H)}$) and after 72 h ($F_{0(72H)}$) were used to calculate the F_0 incremental ratio $(F_{0(72H)} - F_{0(0H)})/F_{0(0H)}$ and to estimate the growth.

The results reported in Figure 5 (bottom panel) are concluded from spots with 0.7 initial OD and show the increase of F_0 after 72 h of different light sources exposure. The histograms indicate that all cyanobacteria strains are capable of growing well under M7 simulated light, with incremental ratio F_0 comparable to those exposed to the solar light. PCC6803, that does not possess the FarLiP gene cluster, has an F_0 incremental ratio close to zero and negligible with respect to the values registered for M7 and solar light. All the other organisms are shown to be capable of exploiting FR light with different extents depending on the strain.

For evaluating the efficiency of the photosynthesis, we analyzed each plate by means of the PAM imaging (FluorCam FC 800MF). We evaluated the PSII photosynthetic efficiency through the chlorophyll fluorescence measurements, before exposing the culture to the various light sources and after 72 h. The photosynthetic efficiency is defined by the ratio F_V/F_m , where $F_V = F_m - F_0$ with F_0 the basal fluorescence, and F_m is the maximum fluorescence assessed after 20 min of dark adaptation followed by a flash of saturating light. The averaged values of the parameters F_V/F_m obtained by the PAM analysis from three independent biological replicates for each organism, derived from the spots with 0.5, 0.7, and 1 initial OD are reported in Table 2. These values show that all the organisms maintain similar photosynthetic efficiency under different light regimes (with values typical of the cyanobacteria strains (Reference [73,75] in Supplementary Materials), while PCC6803 shows a decline of the F_V/F_m parameter under FR light, as expected.

Table 2. Values of F_V/F_m obtained for several organisms. The considered error is 1σ .

Light Source	PCC			
	6803	6912	7203	7335
SOL	0.568 ± 0.050	0.513 ± 0.018	0.501 ± 0.021	0.513 ± 0.022
M7	0.578 ± 0.008	0.501 ± 0.011	0.490 ± 0.026	0.503 ± 0.026
FR	0.380 ± 0.050	0.523 ± 0.028	0.505 ± 0.019	0.560 ± 0.013

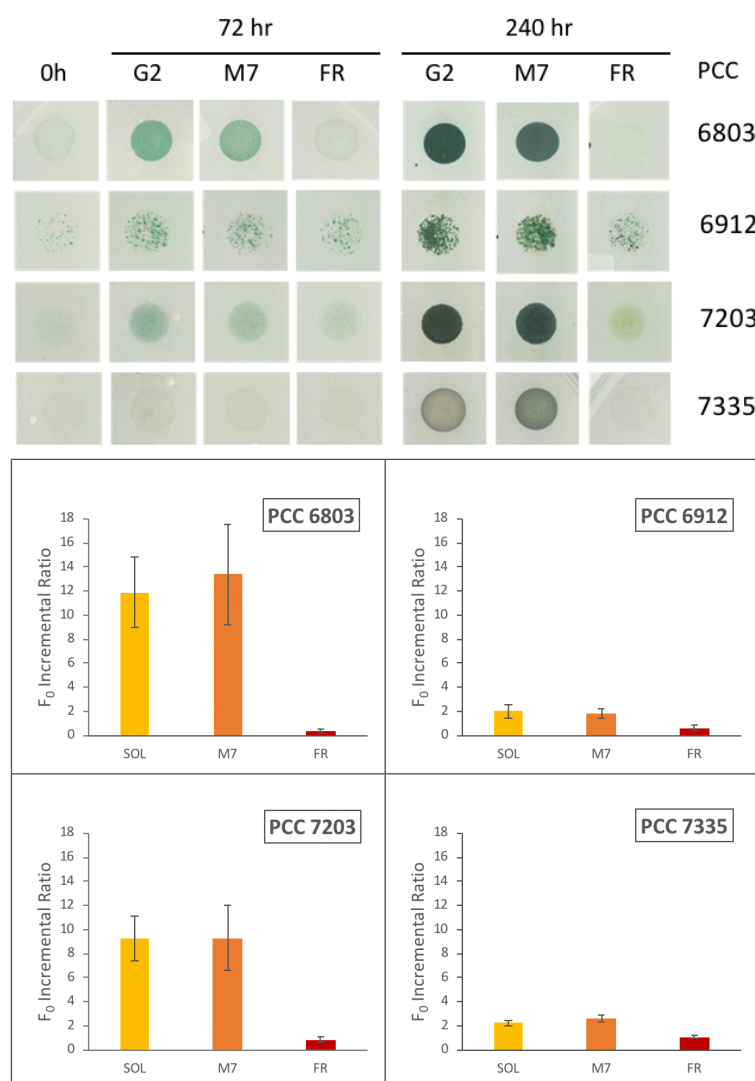


Figure 5. **Top panel:** Photos of the strains in individual plates show the phenotypes of the organisms. From left to right, at the beginning (0 h), after 72 h, and 240 h of exposure to sunlight (SOL), a star M7 (M7), and far-red (FR). The phenotype variation of the spotted strains is related to those with 0.7 initial optical density (OD). **Bottom panel:** Values of the F_0 incremental ratio of the different organisms. The F_0 incremental ratio is defined as follows: $(F_{0(72H)} - F_{0(0H)})/F_{0(0H)}$, with $F_{0(0H)}$ implying F_0 value at 0 h and $F_{0(72H)}$ corresponding to F_0 value at 72 h. The F_0 incremental ratio values are reported in Table S1 in the Supplementary Materials.

5. Discussion and Conclusions

M stars are very popular in the astrobiology community due to their ubiquitous presence in the Galaxy and their small radii, which provide higher amplitude transit signals than solar-like stars. So far, they are recognized to be the most frequent hosts of super-Earths discovered orbiting in the HZ of a star. This sparked off a great theoretical debate about the possibility of having life, particularly photosynthetic life, on these planets. Several efforts have been spent aiming at modeling the upper wavelength limit of putative photoautotrophs on exoplanets. It has been hypothesized that oxygenic photosynthetic organisms could have developed pigments that do not utilize PAR light, but the more abundant NIR light, or employ photosystems using up to 3 or 4 photons per carbon fixed (instead of 2), as well as utilize more photosystems in series (3, 4, or even 6), allowing them to exploit photons of wavelengths up to 2100 nm [21,23,76,77]. Moreover, the prospects for photosynthesis on habitable exomoons via reflected light from the giant planets that they orbit have been theoretically evaluated, suggesting that such photosynthetic biospheres are

potentially sustainable on these moons except those around late-type M-dwarfs [78]. However, up to now, no experimental data (survival, growth, and photosynthetic activity) about the behavior of oxygenic photosynthetic organisms exposed to simulated environmental conditions of exoplanets orbiting the HZ of M dwarfs, in particular, exposed to an M-dwarf light spectrum, have been produced. Numerous investigations (see Reference [67], for a review) have been done, instead, in the field of the oxygenic photosynthesis “beyond 700 nm”, especially after the discoveries of cyanobacteria able to synthesize chl *d* and *f* [79,80]. However, they were committed to understanding the molecular and biochemical mechanisms behind how the photosynthetic functioning of the photosynthetic apparatus under a FR light spectrum, rather than testing it under exotic light spectra [70,71,81–85]. Here, to the best of the authors’ knowledge, for the first time, we present the experimental data obtained directly through exposing photosynthetic organisms to a simulated M dwarf spectrum. We compared the results to responses of those species under solar and FR simulated lights, using innovative laboratory instrumentation. As expected, in FR light, only the cyanobacteria able to synthesize chlorophyll *d* and *f* could grow. Surprisingly, all strains, both able or unable to use FR light, grew and photosynthesized under the M dwarf generated spectrum in a similar way to the solar light and much more efficiently than under the FR one.

In particular, we compared the responses of strains able to have FarLiP and of the control microorganism PCC 6803 that does not. The growth estimated by F_0 incremental ratio parameter obtained for all the cyanobacteria in our study shows a value that is very similar or equal, considering the error bars, to the value of F_0 measured for those spots irradiated with the solar light. In the case of the irradiation with monochromatic light (far-red (FR)), only PCC 6803 is unable to acclimate itself to the FR light, while all the others show a normal photosynthetic efficiency under this light, as well. This suggests that PCC6803 grows very well under simulated M7 light by only using the visible part of the spectrum. The ability of the other organisms to exploit FR light does not seem be beneficial for growth under M7 simulated light. Furthermore, all the tested strains, except PCC 6803, have very similar values of the F_V/F_m under any kind of irradiation spectra. This highlights that they are able to acclimate to all the used lights. Our findings emphasize the importance of simulating both the visible and FR light components of an M dwarf spectrum to correctly evaluate the photosynthetic performances of oxygenic organisms exposed to such an exotic light condition.

Moreover, in a previous work [59], we demonstrated that, with our experimental set up, we can measure the consumption of CO_2 and the production of O_2 of the PCC 6803 cyanobacterium under solar irradiation. This serves as a prelude to the future analysis of the cyanobacteria photosynthetic gas exchanges in real time during their growth under M star spectra irradiation.

Last but not least, we realized an experimental set up that allows us to reproduce, in the laboratory, an alien environment with the possibility to variate the thermal and physical conditions. In this way, we are carrying out experiments on photosynthetic organisms to verify their capacity of thriving and acclimating to extraterrestrial conditions. We developed new and original laboratory devices (e.g., the star irradiation simulator) and novel measurement methods (see Reference [59]) that will allow new experiments in the future. To prepare for the next step of our research plan, we have already produced several models of stable super-Earth atmospheres to be used in the laboratory. We have started to monitor the evolution of oxygen and the fixation of carbon dioxide in the cyanobacteria exposed to very different irradiations and simulated atmospheres.

Hence, if the evolutionary tracks on a habitable planet in the HZ of an M star are quite similar to those on Earth, photosynthetic microorganisms could, as well, produce O_2 and fix CO_2 in organic matter on these planets orbiting such cold stars.

Will it be possible to observe the released oxygen in a remote way? The answer to this question is not simple because it depends not only on the efficiency in producing oxygen by photosynthetic organisms but also on the efficiency of the possible oxygen sinks

that are at work on that planet. The reverse reaction to oxidize photosynthetic products depletes the atmospheric oxygen. The net release of oxygen in the atmosphere, due to this balance, is regulated by the sink of organics in the sediments. If the level of O_2 is low in the atmosphere, the reactions with reducing gases from vulcanism (H_2 and H_2S) and submarine weathering [86,87] can deplete O_2 . If the O_2 production rate is greater than the depletion rate, its build-up in the atmosphere is possible [88], and the Fe^{2+} oxidation process becomes an important one. Catling and Kasting (2017) and Kaltenegger et al. (2010) (Reference [86,87], respectively, and References therein) discussed deeper on the build-up of oxygen in the atmosphere of a planet. Oxygen depletion is a time-dependent process. The atmospheric oxygen is recycled through respiration and photosynthesis in less than 10,000 years. In the case of total extinction of the biosphere of Earth, the atmospheric O_2 would disappear in a few million years [87].

Thus, we conclude that only the observations can give us the right answer. So far, brand new ground- and space-based instruments are planned to be operative with the aim of finding and characterizing extrasolar planets. In the next future, dedicated space missions and space telescopes, like James Webb Space Telescope (JWST) and Origin Space Telescope (OST), and huge ground telescopes will be the right tools to search for life in other worlds.

Supplementary Materials: The following are available online at <https://www.mdpi.com/2075-1729/11/1/10/>. Figure S1: M7 V star input spectrum as it appears before (in light gray), and after (in red) the smoothing process. The emitted spectrum (in blue) is superimposed. The smoothing process reduces the resolution of the input spectra. Hence, the stellar simulator is able to better reproduce the input spectrum following the depletion of flux due to large atomic absorptions or molecular bands. Figure S2: Emitted FR light measured by the FLAME VIS-NIR spectrograph. The central wavelength is 720 nm and the full width at zero level is 130 nm with a wavelength range 650 nm and 780 nm. The luminosity of this lamp is $2.3 \mu\text{mole m}^{-2} \text{s}^{-1}$ in the PAR and $20 \mu\text{mole m}^{-2} \text{s}^{-1}$ in the total working range. Figure S3: Different cultures of the selected cyanobacteria with different optical density, before the 20 μl spots were deposited on the Petri's plate. Figure S4: Examples of a BG-11 agar plate with *S. sp.* PCC6803, *C. fritschii* PCC6912 and *C. thermalis* PCC 7203 spots and a BG-11-ASN III agar plate with *S. sp.* PCC7335. Plates are shown after 72 and 240 h of exposure under the M-dwarf simulated spectrum. Table S1: Averaged values ($n=6$) of the F_0 incremental ratio obtained for several organisms under different light sources. The considered error is 1σ .

Author Contributions: R.C., N.L.R., and L.P. made the conceptualization and supervision of the whole experiment. R.C., M.S.E., L.C., M.B., N.L.R., and L.P. define the methodology necessary to perform the experiment. E.A. and R.C. realized the super-Earths atmospheric models. R.C., L.P., L.C., and M.S.E. conceived and realized the Star Light Simulator (SLS) and the Atmosphere Simulator Chamber (ASC). B.S. realized the control software of the SLS and L.C. the control software of the ASC. B.S., M.S.E., and E.A. calibrated the SLS, M.B., N.L.R., A.C.P., and D.S. conceived the biological experiments. A.C.P., D.S., and M.B. performed them and analyzed the data. L.C., R.C., M.S.E., L.P., and N.L.R. contributed to the analyses. R.C. wrote the manuscript, NLR edited it, and E.A., M.B., L.C., M.S.E., A.C.P., B.S., V.S., L.P. critically read it. All authors have read and agreed to the published version of the manuscript.

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Abbreviations

The following abbreviations are used in this manuscript:

CA	Chromatic Acclimation
CCT	Correlated Color Temperature
CHL	Chlorophyll
COTS	Component Off The Shelf
FaRLiP	Far-red light photoacclimation
HZ	Habitable Zone
JWST	James Webb Space Telescope
NIR	near infra-red
OD	Optical Density
OST	Origin Space Telescope
PAM	Pulse-Amplitude Modulation
PAR	Photochemically Active Radiation
PC	Personal Computer
PCC	Pasteur Culture Collection
TDLAS	Tunable Diode Laser Absorption Spectroscopy
WMS	Wavelength Modulation Spectroscopy

References

1. Dressing, C.D.; Charbonneau, D. The Occurrence Rate of Small Planets around Small Stars. *Astrophys. J.* **2013**, *767*, 95. [\[CrossRef\]](#)
2. Davenport, J.R.A.; Kipping, D.M.; Sasselov, D.; Matthews, J.M.; Cameron, C. MOST Observations of Our Nearest Neighbor: Flares on Proxima Centauri. *Astrophys. J. Lett.* **2016**, *829*, L31. [\[CrossRef\]](#)
3. Mayor, M.; Lovis, C.; Santos, N.C. Doppler spectroscopy as a path to the detection of Earth-like planets. *Nature* **2014**, *513*, 328–335. [\[CrossRef\]](#) [\[PubMed\]](#)
4. Bonfils, X.; Delfosse, X.; Udry, S.; Forveille, T.; Mayor, M.; Perrier, C.; Bouchy, F.; Gillon, M.; Lovis, C.; Pepe, F.; et al. The HARPS search for southern extra-solar planets. XXXI. The M-dwarf sample. *Astron. Astrophys.* **2013**, *549*, A109. [\[CrossRef\]](#)
5. Kopparapu, R.K.; Ramirez, R.; Kasting, J.F.; Eymet, V.; Robinson, T.D.; Mahadevan, S.; Terrien, R.C.; Domagal-Goldman, S.; Meadows, V.; Deshpande, R. Habitable Zones around Main-sequence Stars: New Estimates. *Astrophys. J.* **2013**, *765*, 131. [\[CrossRef\]](#)
6. Seager, S.; Schrenk, M.; Bains, W. An Astrophysical View of Earth-Based Metabolic Biosignature Gases. *Astrobiology* **2012**, *12*, 61–82. [\[CrossRef\]](#)
7. Seager, S.; Bains, W.; Petkowski, J.J. Toward a List of Molecules as Potential Biosignature Gases for the Search for Life on Exoplanets and Applications to Terrestrial Biochemistry. *Astrobiology* **2016**, *16*, 465–485. [\[CrossRef\]](#)
8. Kaltenegger, L. How to Characterize Habitable Worlds and Signs of Life. *Annu. Rev. Astron. Astrophys.* **2017**, *55*, 433–485. [\[CrossRef\]](#)
9. Grenfell, J.L. A review of exoplanetary biosignatures. *Phys. Rep.* **2017**, *713*, 1–17. [\[CrossRef\]](#)
10. Catling, D.C.; Krissansen-Totton, J.; Kiang, N.Y.; Crisp, D.; Robinson, T.D.; DasSarma, S.; Rushby, A.J.; Del Genio, A.; Bains, W.; Domagal-Goldman, S. Exoplanet Biosignatures: A Framework for Their Assessment. *Astrobiology* **2018**, *18*, 709–738. [\[CrossRef\]](#)
11. Schwieterman, E.W.; Kiang, N.Y.; Parenteau, M.N.; Harman, C.E.; DasSarma, S.; Fisher, T.M.; Arney, G.N.; Hartnett, H.E.; Reinhard, C.T.; Olson, S.L.; et al. Exoplanet Biosignatures: A Review of Remotely Detectable Signs of Life. *Astrobiology* **2018**, *18*, 663–708. [\[CrossRef\]](#) [\[PubMed\]](#)
12. Claudi, R.; Alei, E. Biosignatures Search in Habitable Planets. *Galaxies* **2019**, *7*, 82. [\[CrossRef\]](#)
13. Lingam, M.; Loeb, A. Colloquium: Physical constraints for the evolution of life on exoplanets. *Rev. Mod. Phys.* **2019**, *91*, 021002. [\[CrossRef\]](#)
14. Lovelock, J.E. A Physical Basis for Life Detection Experiments. *Nature* **1965**, *207*, 568–570. [\[CrossRef\]](#)
15. Meadows, V.S.; Reinhard, C.T.; Arney, G.N.; Parenteau, M.N.; Schwieterman, E.W.; Domagal-Goldman, S.D.; Lincowski, A.P.; Stapelfeldt, K.R.; Rauer, H.; DasSarma, S.; et al. Exoplanet Biosignatures: Understanding Oxygen as a Biosignature in the Context of Its Environment. *Astrobiology* **2018**, *18*, 630–662. [\[CrossRef\]](#) [\[PubMed\]](#)
16. Lederberg, J. Signs of Life: Criterion-System of Exobiology. *Nature* **1965**, *207*, 9–13. [\[CrossRef\]](#)
17. Seager, S.; Bains, W. The search for signs of life on exoplanets at the interface of chemistry and planetary science. *Sci. Adv.* **2015**, *1*, e1500047. [\[CrossRef\]](#)

18. Harman, C.E.; Schwieterman, E.W.; Schottelkotte, J.C.; Kasting, J.F. Abiotic O₂ Levels on Planets around F, G, K, and M Stars: Possible False Positives for Life? *Astrophys. J.* **2015**, *812*, 137. [\[CrossRef\]](#)
19. Gates, D.M.; Gates, H.J.; Gates, J.C.; Gates, V.R. Spectral properties of plants. *Appl. Opt.* **1965**, *4*, 11. [\[CrossRef\]](#)
20. Gan, F.; Bryant, D. Adaptive and acclimative responses of cyanobacteria to far-red light. *Environ. Microbiol.* **2015**, *17*, 3450–3465. [\[CrossRef\]](#)
21. Wolstencroft, R.D.; Raven, J.A. Photosynthesis: Likelihood of Occurrence and Possibility of Detection on Earth-like Planets. *Icarus* **2002**, *157*, 535–548. [\[CrossRef\]](#)
22. Tinetti, G.; Rashby, S.; Yung, Y.L. Detectability of Red-Edge-shifted Vegetation on Terrestrial Planets Orbiting M Stars. *Astrophys. J. Lett.* **2006**, *644*, L129–L132. [\[CrossRef\]](#)
23. Kiang, N.Y.; Segura, A.; Tinetti, G.; Govindjee; Blankenship, R.E.; Cohen, M.; Siefert, J.; Crisp, D.; Meadows, V.S. Spectral Signatures of Photosynthesis. II. Coevolution with Other Stars And The Atmosphere on Extrasolar Worlds. *Astrobiology* **2007**, *7*, 252–274. [\[CrossRef\]](#) [\[PubMed\]](#)
24. Claudi, R.; Erculiani, M.S.; Galletta, G.; Billi, D.; Pace, E.; Schierano, D.; Giro, E.; D'Alessandro, M. Simulating super earth atmospheres in the laboratory. *Int. J. Astrobiol.* **2016**, *15*, 35–44. [\[CrossRef\]](#)
25. Pecaut, M.J.; Mamajek, E.E. Intrinsic Colors, Temperatures, and Bolometric Corrections of Pre-main-sequence Stars. *Astrophys. J. Suppl.* **2013**, *208*, 9. [\[CrossRef\]](#)
26. Leggett, S.K.; Allard, F.; Dahn, C.; Hauschildt, P.H.; Kerr, T.H.; Rayner, J. Spectral Energy Distributions for Disk and Halo M Dwarfs. *Astrophys. J.* **2000**, *535*, 965–974. [\[CrossRef\]](#)
27. Leggett, S.K.; Allard, F.; Berriman, G.; Dahn, C.C.; Hauschildt, P.H. Infrared Spectra of Low-Mass Stars: Toward a Temperature Scale for Red Dwarfs. *Astrophys. J. Suppl.* **1996**, *104*, 117. [\[CrossRef\]](#)
28. Kasting, J.F.; Whitmire, D.P.; Reynolds, R.T. Habitable Zones around Main Sequence Stars. *Icarus* **1993**, *101*, 108–128. [\[CrossRef\]](#)
29. Selsis, F.; Kasting, J.F.; Levrard, B.; Paillet, J.; Ribas, I.; Delfosse, X. Habitable planets around the star Gliese 581? *Astron. Astrophys.* **2007**, *476*, 1373–1387. [\[CrossRef\]](#)
30. Brown, S.P.; Mead, A.J.; Forgan, D.H.; Raven, J.A.; Cockell, C.S. Photosynthetic potential of planets in 3 : 2 spin-orbit resonances. *Int. J. Astrobiol.* **2014**, *13*, 279–289. [\[CrossRef\]](#)
31. Lingam, M.; Loeb, A. Photosynthesis on habitable planets around low-mass stars. *Mon. Not. R. Astron. Soc.* **2019**, *485*, 5924–5928. [\[CrossRef\]](#)
32. Heath, M.J.; Doyle, L.R.; Joshi, M.M.; Haberle, R.M. Habitability of Planets Around Red Dwarf Stars. *Orig. Life Evol. Biosph.* **1999**, *29*, 405–424. [\[CrossRef\]](#) [\[PubMed\]](#)
33. Lissauer, J.J. Planets Formed in Habitable Zones of M Dwarf Stars Probably Are Deficient in Volatiles. *Astrophys. J. Lett.* **2007**, *660*, L149–L152. [\[CrossRef\]](#)
34. Raymond, S.N.; Scalzo, J.; Meadows, V.S. A Decreased Probability of Habitable Planet Formation around Low-Mass Stars. *Astrophys. J.* **2007**, *669*, 606–614. [\[CrossRef\]](#)
35. Luger, R.; Barnes, R. Extreme Water Loss and Abiotic O₂ Buildup on Planets Throughout the Habitable Zones of M Dwarfs. *Astrobiology* **2015**, *15*, 119–143. [\[CrossRef\]](#)
36. Tian, F.; Ida, S. Water contents of Earth-mass planets around M dwarfs. *Nat. Geosci.* **2015**, *8*, 177–180. [\[CrossRef\]](#)
37. Gillon, M.; Jehin, E.; Lederer, S.M.; Delrez, L.; de Wit, J.; Burdanov, A.; Van Grootel, V.; Burgasser, A.J.; Triaud, A.H.M.J.; Opitom, C.; et al. Temperate Earth-sized planets transiting a nearby ultracool dwarf star. *Nature* **2016**, *533*, 221–224. [\[CrossRef\]](#)
38. Bolmont, E.; Libert, A.S.; Leconte, J.; Selsis, F. Habitability of planets on eccentric orbits: Limits of the mean flux approximation. *Astron. Astrophys.* **2016**, *591*, A106. [\[CrossRef\]](#)
39. Scalzo, J.; Kaltenegger, L.; Segura, A.G.; Fridlund, M.; Ribas, I.; Kulikov, Y.N.; Grenfell, J.L.; Rauer, H.; Odert, P.; Leitzinger, M.; et al. M Stars as Targets for Terrestrial Exoplanet Searches And Biosignature Detection. *Astrobiology* **2007**, *7*, 85–166. doi:10.1089/ast.2006.0125. [\[CrossRef\]](#)
40. Segura, A.; Kasting, J.F.; Meadows, V.; Cohen, M.; Scalzo, J.; Crisp, D.; Butler, R.A.H.; Tinetti, G. Biosignatures from Earth-Like Planets Around M Dwarfs. *Astrobiology* **2005**, *5*, 706–725. doi:10.1089/ast.2005.5.706. [\[CrossRef\]](#)
41. Segura, A.; Walkowicz, L.M.; Meadows, V.; Kasting, J.; Hawley, S. The Effect of a Strong Stellar Flare on the Atmospheric Chemistry of an Earth-like Planet Orbiting an M Dwarf. *Astrobiology* **2010**, *10*, 751–771. [\[CrossRef\]](#) [\[PubMed\]](#)
42. France, K.; Froning, C.S.; Linsky, J.L.; Roberge, A.; Stocke, J.T.; Tian, F.; Bushinsky, R.; Désert, J.M.; Mauas, P.; Vieytes, M.; Walkowicz, L.M. The Ultraviolet Radiation Environment around M dwarf Exoplanet Host Stars. *Astrophys. J.* **2013**, *763*, 149. [\[CrossRef\]](#)
43. Cuntz, M.; Guinan, E.F. About Exobiology: The Case for Dwarf K Stars. *Astrophys. J.* **2016**, *827*, 79. [\[CrossRef\]](#)
44. O'Malley-James, J.T.; Kaltenegger, L. UV Surface Environments of M Star Planets: Surface Habitability and Temporal Biosignatures. In Proceedings of the Habitable Worlds 2017: A System Science Workshop, Laramie, WY, USA, 13–17 November 2017; Volume 2042, p. 4128.
45. O'Malley-James, J.T.; Kaltenegger, L. Biofluorescent Worlds-II. Biological fluorescence induced by stellar UV flares, a new temporal biosignature. *Mon. Not. R. Astron. Soc.* **2019**, *488*, 4530–4545. [\[CrossRef\]](#)
46. Tarter, J.C.; Backus, P.R.; Mancinelli, R.L.; Aurnou, J.M.; Backman, D.E.; Basri, G.S.; Boss, A.P.; Clarke, A.; Deming, D.; Doyle, L.R.; et al. A Reappraisal of The Habitability of Planets around M Dwarf Stars. *Astrobiology* **2007**, *7*, 30–65. [\[CrossRef\]](#) [\[PubMed\]](#)

47. Gale, J.; Wandel, A. The potential of planets orbiting red dwarf stars to support oxygenic photosynthesis and complex life. *Int. J. Astrobiol.* **2017**, *16*, 1–9. [\[CrossRef\]](#)
48. Ranjan, S.; Sasselo, D.D. Influence of the UV Environment on the Synthesis of Prebiotic Molecules. *Astrobiology* **2016**, *16*, 68–88. [\[CrossRef\]](#)
49. Mullan, D.J.; Bais, H.P. Photosynthesis on a Planet Orbiting an M Dwarf: Enhanced Effectiveness during Flares. *Astrophys. J.* **2018**, *865*, 101. [\[CrossRef\]](#)
50. Dressing, C.D.; Charbonneau, D. The Occurrence of Potentially Habitable Planets Orbiting M Dwarfs Estimated from the Full Kepler Dataset and an Empirical Measurement of the Detection Sensitivity. *Astrophys. J.* **2015**, *807*, 45. [\[CrossRef\]](#)
51. Gillon, M.; Triaud, A.H.M.J.; Demory, B.O.; Jehin, E.; Agol, E.; Deck, K.M.; Lederer, S.M.; de Wit, J.; Burdanov, A.; Ingalls, J.G.; et al. Seven temperate terrestrial planets around the nearby ultracool dwarf star TRAPPIST-1. *Nature* **2017**, *542*, 456–460. [\[CrossRef\]](#)
52. Anglada-Escudé, G.; Amado, P.J.; Barnes, J.; Berdiñas, Z.M.; Butler, R.P.; Coleman, G.A.L.; de La Cueva, I.; Dreizler, S.; Endl, M.; Giesers, B.; et al. A terrestrial planet candidate in a temperate orbit around Proxima Centauri. *Nature* **2016**, *536*, 437–440. [\[CrossRef\]](#) [\[PubMed\]](#)
53. Miller-Ricci, E.; Seager, S.; Sasselo, D. The Atmospheric Signatures of Super-Earths: How to Distinguish Between Hydrogen-Rich and Hydrogen-Poor Atmospheres. *Astrophys. J.* **2009**, *690*, 1056–1067. [\[CrossRef\]](#)
54. Petralia, A.; Alei, E.; Aresu, G.; Locci, D.; Cecchi-Pestellini, C.; Micela, G.; Claudi, R.; Ciaravella, A. A systematic study of CO₂ planetary atmospheres and their link to the stellar environment. *Mon. Not. R. Astron. Soc.* **2020**, *496*, 5350–5359. [\[CrossRef\]](#)
55. Bula, R.; Morrow, R.; Tibbitts, T.; Barta, D.; Ignatius, R.; Martin, T. Light-emitting Diodes as a Radiation Source for Plants. *HortScience* **1991**, *26*, 203–205. [\[CrossRef\]](#) [\[PubMed\]](#)
56. Trivellin, N.; Barbisan, D.; Ferretti, M.; Erculiani, M.S.; Claudi, R.U.; Giro, E.; Bonato, M.; Cocola, L.; Poletto, L.; Salasnich, B.; et al. Adaptive multi-wavelength LED star simulator for space life studies. In *Light-Emitting Diodes: Materials, Devices, and Applications for Solid State Lighting XX*; Jeon, H., Tu, L.W., Krames, M.R., Strassburg, M., Eds.; SPIE: Bellingham, WA, USA, 2016; p. 976815. [\[CrossRef\]](#)
57. Salasnich, B.; Claudi, R.U.; Alei, E.; Barbisan, D.; Baruffolo, A.; Cocola, L.; Erculiani, M.S.; La Rocca, N.; Pace, E.; Poletto, L.; et al. Control software for the Multi-Channel Led starlight simulator. In *Software and Cyberinfrastructure for Astronomy V*; SPIE: Bellingham, WA, USA, 2018; Volume 10707, p. 107071I. [\[CrossRef\]](#)
58. Danilović, B.; Cocola, L.; Fedel, M.; Poletto, L.; Savić, D. Formation and Cumulation of CO₂ in the Bottles of the Fermented Milk Drinks. *IPCBE* **2016**, *95*, 26. [\[CrossRef\]](#)
59. Battistuzzi, M.; Cocola, L.; Salasnich, B.; Erculiani, M.S.; Alei, E.; Morosinotto, T.; Claudi, R.; Poletto, L.; La Rocca, N. A New Remote Sensing-Based System for the Monitoring and Analysis of Growth and Gas Exchange Rates of Photosynthetic Microorganisms Under Simulated Non-Terrestrial Conditions. *Front. Plant Sci.* **2020**, *11*, 182. [\[CrossRef\]](#)
60. Erculiani, M.S.; Claudi, R.; Barbisan, D.; Giro, E.; Bonato, M.; Cocola, L.; Farisato, G.; Meneghini, M.; Poletto, L.; Salasnich, B.; et al. A tunable integrated system to simulate colder stellar radiation. In *Optical Systems Design 2015: Optical Design and Engineering VI*; Mazuray, L., Wartmann, R., Wood, A.P., Eds.; SPIE: Bellingham, WA, USA, 2015; Volume 9626, p. 96262D. [\[CrossRef\]](#)
61. Allakhverdiev, S.; Kreslavskii, V.; Zharmukhamedov, S.; Voloshin, R.; Korol'kova, D.; Tomo, T.; Shen, J.R. Chlorophylls *d* and *f* and Their Role in Primary Photosynthetic Processes of Cyanobacteria. *Biochemistry* **2016**, *81*, 201–212. [\[CrossRef\]](#)
62. de Vera, J.P.; Dulai, S.; Kereszturi, A.; Koncz, L.; Lorek, A.; Mohlmann, D.; Marschall, M.; Pocs, T. Results on the survival of cryptobiotic cyanobacteria samples after exposure to Mars-like environmental conditions. *Int. J. Astrobiol.* **2014**, *13*, 35–44. [\[CrossRef\]](#)
63. Billi, D.; Staibano, C.; Verseux, C.; Fagiarone, C.; Mosca, C.; Baqué, M.; Rabbow, E.; Rettberg, P. Dried Biofilms of Desert Strains of *Chroococcidiopsis* Survived Prolonged Exposure to Space and Mars-like Conditions in Low Earth Orbit. *Astrobiology* **2019**, *19*, 1008–1017. [\[CrossRef\]](#)
64. Billi, D.; Baqué, M.; Smith, H.; McKay, C. Cyanobacteria from Extreme Deserts to Space. *Adv. Microbiol.* **2013**, *3*, 80–86. [\[CrossRef\]](#)
65. McKay, C. Thickness of tropical ice and photosynthesis on a snowball earth. *Geophys. Res. Lett.* **2000**, *27*, 2153–2156. [\[CrossRef\]](#) [\[PubMed\]](#)
66. Cockell, C.S.; Raven, J.A. Zones of photosynthetic potential on Mars and the early Earth. *Icarus* **2004**, *169*, 300–310. [\[CrossRef\]](#)
67. Wolf, B.; Blankenship, R. Far-red light acclimation in diverse oxygenic photosynthetic organisms. *Photosynth. Res.* **2019**, *142*, 349–359. [\[CrossRef\]](#) [\[PubMed\]](#)
68. Ains, R.; Temperton, B.; Sambles, C.; Farnham, G.; Skill, S.; Llewellyn, C. Chlorophyll *f* and chlorophyll *d* are produced in the cyanobacterium *Chlorogloeopsis fritschii* when cultured under natural light and near-infrared radiation. *FEBS Lett.* **2014**, *588*, 3770–3777. [\[CrossRef\]](#)
69. Cumbers, J.; Rothschild, L.J. Salt tolerance and polyphyly in the cyanobacterium *Chroococcidiopsis* (Pleurocapsales). *J. Phycol.* **2014**, *50*, 472–482. [\[CrossRef\]](#)
70. Zhao, C.; Gan, F.; Shen, G.; Bryant, D.A. *RfpA*, *RfpB*, and *RfpC* are the Master Control Elements of Far-Red Light Photoacclimation (FaRLiP). *Front. Microbiol.* **2015**, *6*, 1303. [\[CrossRef\]](#)
71. Ho, M.; Gan, F.; Shen, G.; Zhao, C.; Bryant, D.A. Far-red light photoacclimation (FaRLiP) in *Synechococcus* sp. PCC 7335: I. Regulation of FaRLiP gene expression. *Photosynth. Res.* **2017**, *131*, 173–186. [\[CrossRef\]](#)

72. Rippka, R.; Deruelles, J.; Waterbury, J.B.; Herdman, M.; Stanier, R.Y. Generic Assignments, Strain Histories and Properties of Pure Cultures of Cyanobacteria. *Microbiology* **1979**, *111*, 1–61. [\[CrossRef\]](#)
73. Misumi, M.; Katoh, H.; Tomo, T.; Sonoike, K. Relationship between Photochemical Quenching and Non-Photochemical Quenching in Six Species of Cyanobacteria Reveals Species Difference in Redox State and Species Commonality in Energy Dissipation. *Plant Cell Physiol.* **2015**, *57*, 1510–1517. [\[CrossRef\]](#)
74. Perin, G.; Bellan, A.; Segalla, A.; Meneghesso, A.; Alboresi, A.; Morosinotto, T. Generation of random mutants to improve light-use efficiency of *Nannochloropsis gaditana* cultures for biofuel production. *Biotechnol. Biofuels* **2015**, *8*, 161. [\[CrossRef\]](#)
75. Llewellyn, C.A.; Airs, R.L.; Farnham, G.; Greig, C. Synthesis, Regulation and Degradation of Carotenoids Under Low Level UV-B Radiation in the Filamentous Cyanobacterium *Chlorogloeopsis fritschii* PCC 6912. *Front. Microbiol.* **2020**, *11*, 163. [\[CrossRef\]](#) [\[PubMed\]](#)
76. Kiang, N.Y.; Siefert, J.; Govindjee; Blankenship, R.E. Spectral Signatures of Photosynthesis. I. Review of Earth Organisms. *Astrobiology* **2007**, *7*, 222–251. [\[CrossRef\]](#)
77. Takizawa, K.; Minagawa, J.; Tamura, M.; Kusakabe, N.; Narita, N. Red-edge position of habitable exoplanets around M-dwarfs. *Scientific Reports* **2017**, *7*. [\[CrossRef\]](#) [\[PubMed\]](#)
78. Lingam, M.; Loeb, A. Photosynthesis on exoplanets and exomoons from reflected light. *Int. J. Astrobiol.* **2020**, *19*, 210–219. [\[CrossRef\]](#)
79. Miyashita, H.; Ikemoto, H.; Kurano, N. Chlorophyll *d* as a major pigment. *Nature* **1996**, *383*, 402. [\[CrossRef\]](#)
80. Chen, M.; Schliep, M.; Willows, R.D.; Cai, Z.L.; Neilan, B.A.; Scheer, H. A Red-Shifted Chlorophyll. *Science* **2010**, *329*, 1318. [\[CrossRef\]](#)
81. Gan, F.; Zhang, S.; Rockwell, N.C.; Martin, S.S.; Lagarias, J.C.; Bryant, D.A. Extensive remodeling of a cyanobacterial photosynthetic apparatus in far-red light. *Science* **2014**, *345*, 1312–1317. [\[CrossRef\]](#)
82. Ho, M.Y.; Niedzwiedzki, D.M.; MacGregor-Chatwin, C.; Gerstenecker, G.; Hunter, C.N.; Blankenship, R.E.; Bryant, D.A. Extensive remodeling of the photosynthetic apparatus alters energy transfer among photosynthetic complexes when cyanobacteria acclimate to far-red light. *Biochim. Biophys. Acta Bioenerg.* **2020**, *1861*, 148064. [\[CrossRef\]](#)
83. Nürnberg, D.J.; Morton, J.; Santabarbara, S.; Telfer, A.; Joliot, P.; Antonaru, L.A.; Ruban, A.V.; Cardona, T.; Krausz, E.; Boussac, A.; et al. Photochemistry beyond the red limit in chlorophyll *f*-containing photosystems. *Science* **2018**, *360*, 1210–1213. [\[CrossRef\]](#)
84. Ho, M.Y.; Bryant, D.A. Global Transcriptional Profiling of the Cyanobacterium *Chlorogloeopsis fritschii* PCC 9212 in Far-Red Light: Insights Into the Regulation of Chlorophyll *d* Synthesis. *Front. Microbiol.* **2019**, *10*, 465. [\[CrossRef\]](#)
85. Kurashov, V.; Ho, M.Y.; Shen, G.; Piedl, K.; Laremore, T.N.; Bryant, D.A.; Golbeck, J.H. Energy transfer from chlorophyll *f* to the trapping center in naturally occurring and engineered Photosystem I complexes. *Photosynth. Res.* **2019**, *141*, 151–163. [\[CrossRef\]](#)
86. Catling, D.C.; Kasting, J.F. *Atmospheric Evolution on Inhabited and Lifeless Worlds*; Cambridge University Press: Cambridge, UK, 2017.
87. Kaltenegger, L.; Selsis, F.; Fridlund, M.; Lammer, H.; Beichman, C.; Danchi, W.; Eiroa, C.; Henning, T.; Herbst, T.; Léger, A.; et al. Deciphering Spectral Fingerprints of Habitable Exoplanets. *Astrobiology* **2010**, *10*, 89–102. [\[CrossRef\]](#) [\[PubMed\]](#)
88. Lehmer, O.R.; Catling, D.C.; Parenteau, M.N.; Hoehler, T.M. The Productivity of Oxygenic Photosynthesis around Cool, M Dwarf Stars. *Astrophys. J.* **2018**, *859*, 171. [\[CrossRef\]](#)